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Perkin 1 Abstracts: Biocatalysis in Organic Synthesis aims to cover recent literature concerning the applications of enzymes and micro-organisms as catalysts in organic synthesis. The abstracts will emphasise the key synthetic step(s) that are mediated by the biocatalyst. Emerging technologies for biocatalyst design and optimisation will also be included.

Biocatalytic route to well-defined macromers built around a sugar core HO vinyl methacrylate, THF, 30°C, 8h, AmanolPS 8h, Novozyme-435 vinyl methacrylate, THF 30°C, 8h, Novozyme-435 $R = -COC(Me) = CH_2$ $R_1 = H$ Selective acrylation of 4-C-hydroxymethyl-1,2-O-isopropylidene-α-D-pentofuranose or by Amano PS occurred at the C-5 hydroxy and C-1' position with Novozyme-435. R = HThe acryl sugar was ring opened and polymerised in toluene with Novozyme-435 $R_1 = -COC(Me) = CH_2$ with the acryl sugar moiety linked to the carboxy terminus of a PCL chain. This was derivatised with oxalyl chloride which showed that <5% of the consumed ε-CL R. Kumar and R. A. Gross, J. Am. Chem. Soc., 2002, 124, 1850 formed chains with a carboxy terminus.

The effect of polyphenols on the enantioselectivity of lipase-catalysed reactions

Lipase

OCOPh
Pseudomonas lipase Amano AH
phosphate buffer pH 7.0
30°C, 96h
Without additive
37% conversion
41% e.e.
69% e.e.
$$E = 8$$
In the presence of
 $E = 8$
 $E = 8$

K. Nakamura and K. Takenaka, Tetrahedron: Asymmetry, 2002, 13, 415.

Various catechin derivatives were screened and found to enhance the stereoselectivity of the hydrolysis of 1b using lipase. In addition, some simple phenol derivatives were also found to increase the enantioselectivity

Chemo-enzymatic synthesis of (1S,5R)-(-)-frontalin

Lipase

R. Chênevert and D. Caron, Tetrahedron: Asymmetry, 2002, 13, 339

S. Conde and P. López-Serrano, Eur. J. Org. Chem., 2002, 922

(R)-2 was chemically transformed into frontalin in 95% yield.

Amidation of N-blocked L- and D-aspartic acid diesters Lipase Candida antarctica lipase B CO₂Et PG-HN CO-NH-Pn CO-NH-Pn CO₂Et PG-HN PG-HN PG-HN. diisopropyl ether CO-NH-Pn CO₂Et CO₂Et CO-NH-Pn ò n-pentyl (Pn) amine 1a-d 4a-d 2a-d PG = Protecting Group Possible products a = CBz; b = Boc; c = Ac; d = PhAcDerivatives of aspartic acid 1a-d were treated with n-pentylamine in the presence of lipase B from Candida antarctica to yield a mixture of the four possible products shown. L- derivatives underwent amidation at the α -position (2a-d) preferentially. D- N- amide derivatives were

converted to β-monoamides (3c and 3d).

Enantioselective lipase catalysed reactions of methyl pipecolinate

CAL-B, BuOH,
iPrCO₂H, 82%

CO₂Me

(±) 1

CAL-A, Et₂O,
iPrCO₂H, 46%

A. Liljeblad, J. Lindborg, A. Kanerva, J. Katajisto and L.T. Kanerva, *Tetrahedron Lett.*, 2002, **43**, 2471.

A study of the use of $Candida\ antarctica\$ lipase in the resolution of N-pipecolinate esters is reported. CAL-A was found to be a highly enantio- and chemoselective catalyst for the acylation of secondary amine, 1, whereas CAL-B reacted chemoselectively at the ester function under the same conditions.

Kinetic resolution and fluorous phase labeling.

Lipasa

Lipase

B. Hungerhoff, H. Sonnenschein and F. Theil, *J. Org. Chem.*, 2002, **67**, 1781.

The highly fluorinated ester 1 was synthesised in 5 steps and used as the acyl donor in a lipase catalysed resolution to yield ester 2. The product mix was partitioned between n-C₆F₁₄ and methanol. 3 other alcohols were effectively resolved in this way.

Nonaqueous biocatalytic synthesis of new cytotoxic doxorubicin derivatives: exploiting unexpected differences in the regioselectivity of salt-activated and solubilised subtilisin

O O OH O OH OH

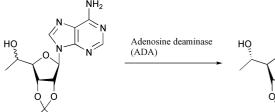
co-lyophilised subtilisin Carlsberg

D. H. Altreuter, J. S. Dordick and D. S. Clark, *J. Am. Chem. Soc.*, 2002, **124**, 1871.

The nonaqueous acylation of doxorubicin was catalysed with ion-paired subtilisin Carlsberg with aerosol OT. Co-lyophilised subtilisin Carlsberg with 98% (w/w) KCl expanded the enzyme's regiospecificity to enable addition acylation at C4' hydroxy and C3' amine groups.

Preparation of diastereomerically pure 5'-methyl adenosine and inosine derivatives $$\operatorname{NH}_2$$

NH₂



A study of the use of ADA to catalyse the deamination of adenosine analogs containing the ribose moiety modified at positions 2', 3' and 5', is reported. This process allows the preparation of diastereomerically pure (5'S)-inosine and unreacted (5'R)-adenosine derivatives.

P. Ciuffreda, A. Loseto and E. Santaniello, *Tetrahedron*: *Asymmetry*, 2002, **13**, 239.

Characterisation of Thermotoga maritima KDPG aldolase

Aldolase

Adenosine

deaminase

J. S. Griffiths, N. J. Wymer, E. Njolito, S. Niranjanakumari, C. A. Fierke and E.J. Toone, *Bioorg. Med. Chem.*, 2002, 10, 545.

The KDPG aldolase from T. maritima has been cloned and over-expressed in E. coli. The scheme shown was used to establish stereospecificty of the enzyme. The activity of the retro-aldol cleavage was also studied. Substrate specificity in the synthetic direction was studied for a variety of aldehydes.

Remote chiral recognition in a reaction mediated by a catalytic antibody

Antibody

$$O_2N$$
 O_2N
 O_2N

R'= H or Me

L. J. D'Souza, B. Gigant, M. Knossow and B. S. Green, J. Am. Chem. Soc., 2002, 124, 2114.

D2.3 antibody preferentially binds the L-enantiomer of the chiral 4-nitrobenzyl ester hapten. The antibody catalyses its hydrolysis with a 20-fold improvement by a modification at the binding site from a glycine to an alanine.

Antibody-catalysed regiocontrolled aldol reactions

 $1, X = CH_2, R = Me$

4, X = S, R = Me

2, $X = CH_2$, R = Et3, X = 0, R = Me

OH

5, $X = CH_2$, R = Me

6, $X = CH_2$, R = Et

7, X = O, R = Me

8, X = S, R = Me

9, $X = CH_2$, R = Me

ОН

10, $X = CH_2$, R = Et

11, X = O, R = Me

12, X = S, R = Me

V. Maggiotti, M. Resmini and V. Gouverneur, Angew. Chem., Int. Ed., 2002, 41, 1012.

Antibody Ab84G3 catalysed the regioselective aldol coupling of benzaldehyde to unsymmetrical ketones 1 to 4 to yield products of reaction at the less substituted carbon atom 5-8. 9 to 12 were not formed. The enantioselectivity of addition was typically higher than 94%.

Synthesis of a bifunctionalised cellohexaoside intermediate

Cellulase

Antibody 84G3

V. Bover, S. Fort, T.P. Frandsen, M. Schülein, S. Cottaz and H. Driguez, Chem. Eur. J., 2002, 8, 1389.

transglycosylation of α-glucosylfluoride The was achieved the Glu197Ala 'glycosynthase' mutant of cellulase Cel7B from Humicola insolens. Condensation of 1 with an equimolar amount of cellobiose, followed by acetylation gave tetrasaccharide 2 with a yield of 66%.

First asymmetric epoxidation catalysed by cyclohexanone monooxygenase

Cyclohexanone monooxygenase

S. Colonna, N. Gaggero, G. Carrea, G. Ottolina, P. Pasta and F. Zambianchi, Tetrahedron Lett., 2002, 43, 1797.

The first highly enantioselective epoxidation reaction catalysed by cyclohexanone monooxygenase is reported. Diethyl vinylphosphonate, 1, was converted to diethyl phosphonate epoxide, 2, in moderate yield and excellent e.e.

Enantioselective microbial reduction of an aryl ketone

Dehydrogenases

R. N. Patel, L. Chu, R. Chidambaram, J. Zhu and J. Kant, Tetrahedron: Asymmetry, 2002, 13, 349.

(R)-2 is an intermediate in the synthesis of (R)-3-fluoro-4-[[hydroxy(5,6,7,8tetrahydro-5,5,8,8-tetramethyl-2-naphthyl)acetyl]amino]benzoic acid, retinoic acid receptor gamma-specific agonist which is potentially useful as a dermatological and anti-cancer drug.

A computer program for the determination of stereoselectivity in biocatalytic enantioconvergent reactions

inversion at C2 + H₂O ÓН (2R, 3S)

Epoxide Hydrolases/ Dehalogenases/Sulfatases/ Glycosidases

retention at C2
retention at C2
$$\begin{array}{c} k_1 \\ k_2 \\ (2S,3S) \end{array}$$

$$+ H_2O \xrightarrow{\text{inversion at C2}} k_3 \xrightarrow{\text{inversion at C2}} OH \\ (2S,3R) \\ OH \\ (2R,3R) \end{array}$$

K. Faber and W. Kroutil, Tetrahedron: Asymmetry, 2002, 13, 377.

A parameter 'RI' (Retention-Inversion ratio) was introduced to describe the stereoselectivity of biocatalytic asymmetric reactions which may proceed via different regio- or stereo-isomeric pathways. A computer program was developed for the treatment of the kinetics of such single-step processes.

Biosynthesis of an organofluorine molecule

Fluorinase

D. O'Hagan, C. Schaffrath, S. L. Cobb, J. T. G. Hamilton and C. D. Murphy, Nature, 2002, 416, 279.

NH_2 ΗÓ OH

8 steps

(2S, 3R, 4S)-4-hydroxyleucine 3.

A partially purified cell extract from Streptomyces cattleya was able to transform Sadenosylmethionine (SAM) 1 to fluoroacetate 3 when supplemented with fluoride ions. The 'fluorinase' appeared to catalyse a nucleophilic attack at C-5' of SAM to give 2. 2 may be metabolised to 3 via fluoroacetaldehyde.

Preparation of a precursor of (2S, 3R, 4S)-4-hydroxyleucine

Geotrichum candidum

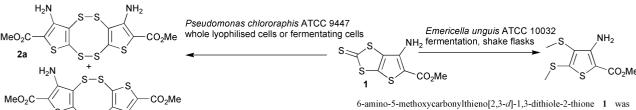


Q. Wang, J. Ouazzani, N. André-Sasaki and P. Potier, Eur. J. Org. Chem., 2002, 834.

Ethyl 2-methylacetoacetate 1 was reduced by resting cells of Geotrichum candidum LCM I-2366 to give (2S, 3S)-3-hydroxy-2-methylbutanoate 2 in 91% yield with 97% diastereomeric excess and 92% enantiomeric excess. Eight further synthetic steps gave the insulinotropic α-amino acid

First preparative biocatalytic hydrolysis and Smethylation of cyclic trithiocarbonates

Microbial



W. Kroutil, A. A. Stämpfli, R. Dahinden, M. Jörg, U. Müller and P. Pachlatko, Tetrahedron, 2002, 58, 2589.

hydrolysed by *Pseudomonas chlororaphis* ATCC 9447 and oxidatively dimerised to form tetrathiocin derivatives 2a and 2b. The cyclic trithiocarbonate was also methylated by Emericella unguis ATCC 10032 involving cleavage followed by methylation of both thiol groups

Butane and propane oxidation

2h

Monooxygenase

Using mutants of $P450_{cam}$, in which the active site volume was reduced, the oxidation of butane and propane was studied. Turnover rate and efficiency generally increased with decreasing active site volume. The highest turnover rate (750 min⁻¹) was observed for butane using a quadruple S. G. Bell, J.-A. Stevenson, H. D. Boyd, S. Campbell, A. D. Riddle, mutant. This value is comparable to the camphor oxidation rate by the wild-type enzyme (1200 E. L. Orton and L.-L. Wong, *Chem. Commun.*, 2002, 490.

Parallel kinetic resolution of β-keto nitriles

(±)

M. isabellina
NRRL 1757

M. isabellina
NRRL 1757

OH

yield 54%
e.e. 74%

OH

yield 38%
e.e. 99%

(R)

ОН

After separating the alcohol diastereomers by chromatography, oxidation gave the (R)- and (S)-ketones. Enhancement of the e.e. of the / alcohol was achieved by a repeat of the reduction using the (S)-ketone, giving J. R. Dehli and V. Gotor, J. Org. Chem., 2002, 67, 1716. the alcohol in 44% yield and 96% e.e. Similar results were obtained for the analogous 6 membered ketones.

Coupling of pyrimidine bases to 2-deoxy-D-ribose-5-phosphate

H₂O₃PO OH phosphoribomutase OH OPO₃H₂

N. Ouwerkerk, M. Steenweg, M. de Ruijter, J. Brouwer, J. H. van Boom, J. Lugtenburg and J. Raap, *J. Org. Chem.*, 2002, **67**, 1480.

thymidine phosphorylase (TP) thymidine phosphorylase OH thymidine HO ON N OH Thymidine HO ON OH Thymidine H

The base coupling was carried out in two enzymatic steps from 2-deoxy-D-ribose-5-phosphate. TP shows no activity with cytosine and 4 chemical steps were used to convert 2'-deoxyuridine to 2'-deoxycytidine. It was possible using this route to synthesise a variety of labeled pyrimidine 2'-deoxynucleosides.

Protease-mediated fragmentation of *p*-aminobenzyl ethers: a new strategy for the activation of anticancer prodrugs

Protease

deoxyuridine

Mortierella isabellina

Phosphoribomutase,

Peptide
$$N$$
 $X = NH, O$ N N NH, O $NH,$

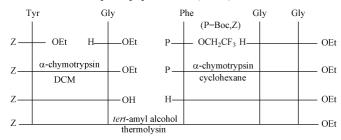
uracil

B. E. Toki, C. G. Cerveny, A. F. Wahl and P. D. Senter, *J. Org. Chem.*, 2002, **67**, 1866.

N-protected peptide benzyloxycarbonyl-valine-citrulline (Z-val-cit), was attached to amino groups of p-amidobenzyl ether derivatives of 1-naphthol and N-acetylnorephedrine. Upon treatment with cathepsin B peptide hydrolysis occured with 1-naphthol release. On the basis of these results etoposide and combretastatin were attached to Z-val-cit-p-amidobenzyl alcohol through ether linkages. Thus forming peptide drug derivatives that were stable in aqueous buffer and serum and underwent ether fragmentation upon treatment with cathepsin B.

Synthesis of a precursor of bioactive pentapeptide OGP(10-14)

Protease



P. Liu, G. Tian, K.-S. Lee, M.-S. Wong and Y. Ye, *Tetrahedron Lett.*, 2002, **43**, 2423.

The enzymatic synthesis of a precursor of the bioactive pentapeptide OGP(10-14) using proteases in organic solvents is described. The effect of different solvents, water content and addition of triethylamine on the reaction was also investigated.

A facile 1,5-rearrangement of β-formylenamides

Pseudomonas fluorescens

R = H, COMe, COPh

U. Bora, M. Longchar, A. Chetia, B. S. D. Kumar, R. C. Boruah and J. S. Sandhu, *Tetrahedron Lett.*, 2002, **43**, 2269.

The use of *Pseudomonas fluorescens* strain RRLJ 134 to catalyse the 1,5-rearrangement of β -formylenamides to the corresponding β -acetoxyenones is reported. This microbe was also found to be an efficient catalyst for the cleavage of acetates.

Chemo-enzymatic synthesis of (2R)-8-substituted-2-aminotetralins

Pseudomonas fluorescens

Pseudomonas fluorescens N3 recombinant strain

30°C, 1-24h

b) R = C₂H₅ c) R = OCH₃ d) R = COOCH₃ e) R = Cl

f) $R = NO_2$

F. Orsini, G. Sello, E. Travaini and P. Di Gennaro, *Tetrahedron: Asymmetry*, 2001, **13**, 253.

Substrate **2d** is a precursor in the synthesis of (2R)-2-amino-8-methoxy-1,2,3,4-tetrahydronaphthalene.

A solvent stable alcohol dehydrogenase activity

W. Stampfer, B. Kosjek, C. Moitzi, W. Kroutil and K. Faber, *Angew. Chem., Int. Ed.*, 2002, **41**, 1014.

Rhodococcus ruber

1, R = -(CH₂)₂-CH=CMe₂
2, R =
$$n$$
-C₆H₁₃
3, R = Ph

Lyophilised cells of *Rhodococcus ruber* DSM 44541 were reported to be a highly efficient catalyst for the preparative reduction/oxidation of ketones/alcohols in the presence of high concentrations of substrate and co-substrate for cofactor regeneration. Ketone **1** (125 gL⁻¹) was reduced to the corresponding (*S*)-alcohol with 99% e.e. in phosphate buffer containing 50% propanol.

Biosynthesis of novel spinosyns

$$\begin{array}{c} R_2 \\ O \\ O \\ O \\ H \\ H \\ H \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ \end{array}$$

$$\begin{array}{c} O \\ \end{array}$$

$$\begin{array}{c}$$

S. Gaisser, C. J. Martin, B. Wilkinson, R. M. Sheridan, R. E. Lill, A. J. Weston, S. J. Ready, C. Waldron, G. D. Crouse, P. F. Leadlay and J. Staunton, *Chem. Commun.*, 2002, 618.

Saccharopolyspora erythraea

3.
$$R_1 = H$$
, $R_2 = HO$
4. $R_1 = CH_3$, $R_2 = HO$
5. $R_1 = H$, $R_2 = HO$
6. $R_1 = CH_3$, $R_2 = HO$
HO
HO
HO
HO
HO
HO
HO
HO

The novel spinosyns (3-6) were obtained on feeding a mix of 1 and 2 to the mutant *Saccharopolyspora erythaea* strain SGT2pSGSpnP.